

LIPOPUREALINS D AND E AND PUREALIDIN H, NEW BROMOTYROSINE ALKALOIDS FROM THE OKINAWAN MARINE SPONGE *PSAMMAPLYSILLA PUREA*

JUN'ICHI KOBAYASHI,* KAORI HONMA, MASASHI TSUDA,

Faculty of Pharmaceutical Sciences, Hokkaido University, Sapporo 060, Japan

and TOSHIYUKI KOSAKA

Analytical and Metabolic Laboratories, San'kyo Co., Ltd., Tokyo 140, Japan

ABSTRACT.—Three new bromotyrosine alkaloids, lipopurealins D and E, and purealidin H, have been isolated from the Okinawan marine sponge *Psammaphysilla purea* and the structures elucidated on the basis of 2D nmr and fabms/ms data.

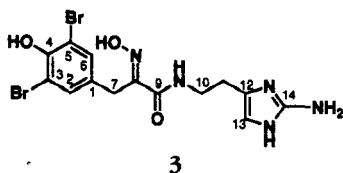
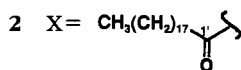
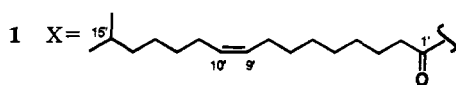
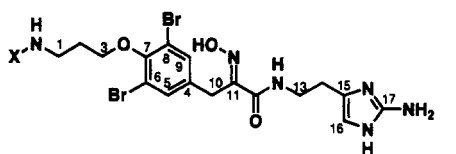
Marine sponges belonging to the order Verongidae have proven to be a rich source of bromotyrosine alkaloids (1), among which lipopurealins A–C, isolated from the Okinawan marine sponge *Psammaphysilla purea* Carter (Aplysinellidae) (2) are unique metabolites with a long acyl chain. In our search for bioactive substances from Okinawan marine organisms (3–5), extracts of this sponge were further examined to obtain two new lipopurealin congeners, lipopurealins D [1] and E [2], and a new bromotyrosine derivative, purealidin H [3]. We describe herein the isolation and structure elucidation of 1–3.

The EtOAc-soluble fraction of the MeOH extract of this sponge collected

off the Kerama Islands, Okinawa, Japan, was subjected to Si gel and Sephadex LH-20 cc and C₁₈ hplc to afford lipopurealins D [1] (0.0004%, wet wt) and E [2] (0.0007%), and purealidin H [3] (0.0004%), together with the known related compounds, lipopurealin B (2) and purealin (6).

Hrfabms data of compounds 1 and 2 established the molecular formulas C₃₄H₅₂N₆O₄Br₂ (*m/z* 767.2540, M⁺+H, Δ +4.5 mmu) and C₃₆H₅₈N₆O₄Br₂ (*m/z* 797.2956, M⁺+H, Δ -0.8 mmu), respectively. Ir absorptions at 3400 and 1675 cm⁻¹ were attributed to NH/OH and amide carbonyl groups, respectively. The uv spectra of 1 and 2 showed the characteristic absorption (λ max 284 nm) of substituted aromatic ring(s). The ¹H- and ¹³C-nmr features of 1 and 2 suggested that these compounds contain a common bromotyrosine unit (C-1–C-17) with an aminohistamine moiety, corresponding to C₁₇H₂₁N₆O₃Br₂, and a long acyl chain, C₁₇H₃₁O and C₁₉H₃₇O, respectively. The ¹³C-nmr chemical shifts of C-11 (δ_C 152.9) suggested that C-11 was assignable to the carbon of an α-ketoxime (7). The *E*- geometry of the oxime at C-11 of 1 and 2 was deduced from the chemical shift of C-10 (δ_C 28.9) (7).

The ¹H-nmr spectrum of 1 revealed signals due to a doublet methyl (δ_H 0.86, 6H), a multiplet methine (δ_H 1.51), and a disubstituted olefin (δ_H 5.33, 2H), im-



plying the presence of an unsaturated long acyl chain with a branched terminus. The position of the double bond was revealed from daughter ion peaks observed in the collisionally activated dissociation (cad) (8) mass spectrum for a fragment ion (m/z 308), corresponding to **A** (Figure 1). The intense fragment ions characteristic for two allyl positions at m/z 168 and 222 and a homoallyl position at m/z 236 indicated that the double bond was located at C-9' (9). *Z*-Geometry of the olefin was deduced from the chemical shift (δ_C 27.9) of the allylic carbons (C-8' and C-11) (10). Thus, the structure of lipopurealin D was elucidated as **1**.

The ^1H -nmr spectrum of lipopurealin E [**2**], containing signals due to a triplet methyl [δ_H 0.88, 3H] and a long acyl chain (30H, δ_H 1.33–1.26), indicated the presence of a *n*-nonadecanoyl group at C-1. Thus, the structure of lipopurealin E was assigned as **2**.

The molecular formula, $\text{C}_{14}\text{H}_{15}\text{N}_5\text{O}_3\text{Br}_2$, of purealidin H [**3**] was established by

hrfabms (m/z 459.9631, $\text{M}^+ + \text{H}$, $\Delta + 1.1$ mmu). The ^1H - and ^{13}C -nmr spectra of **3** revealed all the resonances corresponding to C-4 through C-17 of lipopurealins D [**1**] and E [**2**] except for a phenolic hydroxy proton signal (δ_H 9.77). The structure of purealidin H was thus determined as **3**.

Lipopurealins D [**1**] and E [**2**] isolated from the sponge *Psammaphysilla purea* are new lipopurealin congeners with a long acyl chain. Similar bromotyrosine alkaloids with a long acyl chain have been reported in marine sponges as follows: lipopurealins A–C (**2**) from *P. purea*, araplysillin 2 (**11**) from *P. arabica*, psammaphysin D (**12**) from *Aplysinella* sp., and aplysamine 5 (**13**) from *P. purpurea*. Lipopurealins D [**1**] and E [**2**] and purealidin H [**3**] showed no cytotoxicity (>10 $\mu\text{g/ml}$).

EXPERIMENTAL

GENERAL EXPERIMENTAL METHODS.—Uv and ir spectra were taken on Jasco Ubest-35 and Jasco ir Report-100 spectrometers, respectively. ^1H -

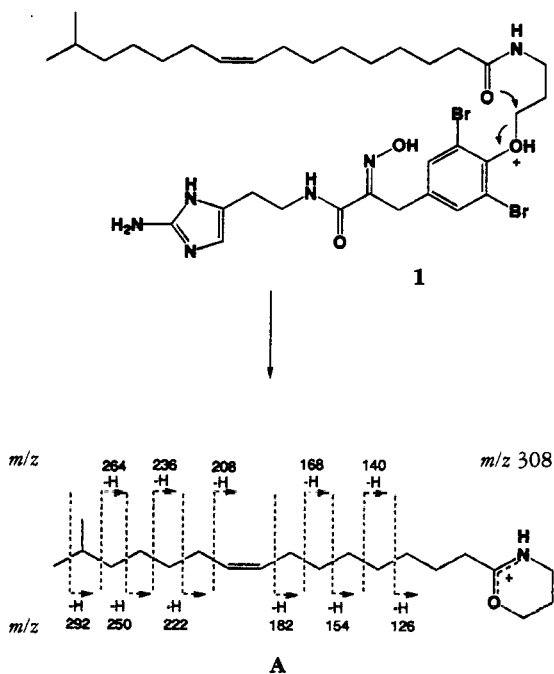


FIGURE 1. Fabms/ms fragmentations of m/z 308 from lipopurealin D [**1**].

and ^{13}C -nmr spectra were conducted with JEOL EX-400 and GSX-270 spectrometers. Fabms and fabms/ms spectra were recorded employing JEOL HX-110 and VG70-4SE spectrometers, respectively.

ANIMAL MATERIAL.—The dark-brown sponge, *Psammaphysilla purea* Carter, was collected off the Kerama Islands, Okinawa, Japan, and kept frozen until used. A voucher specimen is deposited at the Mukaishima Marine Biological Station.

EXTRACTION AND ISOLATION.—The sponge (1.0 kg, wet wt) was extracted with MeOH (1.3 liters \times 2). After evaporation under reduced pressure, the residue (44.1 g) was partitioned between EtOAc (400 ml \times 3) and 1 M aqueous NaCl. A portion (1.07 g) of the EtOAc-soluble material was subjected to Si gel cc [CHCl₃-*n*-BuOH-HOAc-H₂O (1.5:6:1:1)] and then passage over a Sephadex LH-20 column [CHCl₃-MeOH (1:1)] to give purealin (0.018%, wet wt), crude purealidin H, and a mixture of lipopurealins. The mixture of lipopurealins was purified by C₁₈ hplc [YMC Pack AM-323 ODS, 10 \times 250 mm; flow rate, 2.0 ml/min; uv detection at 230 nm] with MeOH-H₂O (9:1) containing 0.2 M aqueous NaCl to afford lipopurealins D [1] (0.0004%, wet wt, *R*, 31 min), E [2] (0.0007%, *R*, 76 min), and B (0.0014%, *R*, 28.5 min). Purealidin H [3] (0.0004%, *R*, 6.5 min) was obtained by C₁₈ hplc [YMC Pack AM-323 ODS, 10 \times 250 mm; eluent CH₃CN-H₂O-TFA (35:65:0.1); flow rate, 2.0 ml/min; uv detection at 230 nm].

Lipopurealin D [1].—Colorless amorphous solid: uv (MeOH) λ max 284 (ϵ 1100), 274 nm (1400); ir (KBr) ν max 3400, 2910, 1675, 1625, 1535, 1450, 1250, 1120 cm⁻¹; ^1H nmr (CD₃OD, 400 MHz) δ 7.46 (2H, s, H-5 and H-9), 6.41 (1H, s, H-16), 5.33 (2H, t, *J*=5.4 Hz, H-9' and H-10'), 4.00 (2H, t, *J*=6.1 Hz, H₂-3), 3.82 (2H, s, H₂-10), 3.45 (2H, t, *J*=6.8 Hz, H₂-13), 3.43 (2H, t, *J*=7.3 Hz, H₂-1), 2.66 (2H, t, *J*=6.8 Hz, H₂-14), 2.18 (2H, t, *J*=7.3 Hz, H₂-2'), 2.03 (6H, m, H₂-2), H₂-8', and H₂-11'), 1.60 (2H, m, H₂-3'), 1.51 (1H, m, *J*=6.8 Hz, H-15'), 1.35–1.27 (14H, m, CH₂), 0.86 (6H, d, *J*=6.8 Hz, H₃-16', and H₃-17'); ^{13}C nmr (CD₃OD, 100 MHz) δ 177.2 (s, C-1'), 166.3 (s, C-12), 153.7 (s, C-7), 152.9 (s, C-11), 148.3 (s, C-17), 138.2 (s, C-4), 135.3 (2C, d, C-5 and C-9), 131.7 (d, C-9'), 131.6 (d, C-10'), 126.2 (s, C-15), 119.6 (2C, s, C-6 and C-8), 112.0 (d, C-16), 73.1 (t, C-3), 40.2 (t, C-13), 38.5 (t, C-1), 38.0 (t, C-2'), 31.9–31.1 (8C, t), 31.1 (t, C-2), 30.0 (d, C-15'), 28.9 (t, C-10), 27.9 (t, 2C, C-8' and C-11'), 27.4 (t, C-3'), 24.5 (t, C-14), 23.8 (2C, q, C-16' and C-17'); fabms *m/z* [M+H]⁺ 767, 769, 771 (1:2:1); hrfabms found *m/z* 767.2540 calcd for C₃₄H₃₃N₆O₄Br₂ [M+H]⁺ 767.2495.

Lipopurealin E [2].—Colorless amorphous solid: uv (MeOH) λ max 284 (ϵ 710), 274 nm

(1000); ir (KBr) ν max 3400, 2840, 1675, 1620, 1535, 1450, 1200, 1130 cm⁻¹; ^1H nmr (CD₃OD, 400 MHz) δ 7.46 (2H, s, H-5 and H-9), 6.46 (1H, s, H-16), 4.00 (2H, t, *J*=6.1 Hz, H₂-3), 3.82 (2H, s, H₂-10), 3.45 (2H, t, *J*=6.8 Hz, H₂-13), 3.43 (2H, t, *J*=6.8 Hz, H₂-1), 2.67 (2H, t, *J*=6.8 Hz, H₂-14), 2.18 (2H, t, *J*=7.6 Hz, H₂-2'), 2.03 (2H, t, *J*=6.1 and 6.8 Hz, H₂-2), 1.60 (2H, m, H₂-3'), 1.33–1.26 (30H, m, CH₂), 0.89 (3H, t, *J*=6.8 Hz, H₃-19'); ^{13}C nmr (CD₃OD, 100 MHz) δ 177.2 (s, C-1'), 166.4 (s, C-12), 153.6 (s, C-7), 152.9 (s, C-11), 148.5 (s, C-17), 138.1 (s, C-4), 135.3 (2C, d, C-5 and C-9), 126.4 (s, C-15), 119.6 (2C, s, C-6 and C-8), 111.7 (d, C-16), 73.1 (t, C-3), 39.9 (t, C-13), 39.0 (t, C-1), 38.0 (t, C-2'), 34.7–31.2 (14C, t), 31.1 (t, C-2), 29.6 (t, C-17'), 28.9 (t, C-10), 27.9 (t, C-18'), 24.5 (t, C-14), 15.2 (q, C-19'); fabms [M+H]⁺ *m/z* 797, 799, 801 (1:2:1); hrfabms found *m/z* 797.2956, calcd for C₃₆H₃₉N₆O₄Br₂ [M+H]⁺ 797.2964.

Purealidin H [3].—Colorless oil; uv (MeOH) λ max 284 (ϵ 1400), 277 nm (1700); ir (KBr) ν max 3400, 2920, 2845, 1680, 1520, 1470, 1200, 1135, 1120 cm⁻¹; ^1H nmr (DMSO-*d*₆, 400 MHz) δ 11.98 (1H, br s, 13-NH), 11.93 (1H, s, 8-NOH), 11.56 (1H, br s, 12-NH), 9.77 (1H, br s, 4-OH), 8.13 (1H, t, *J*=5.9 Hz, NH-9), 7.33 (2H, s, 14-NH₂), 7.32 (2H, s, H-2 and H-6), 6.57 (1H, s, H-13), 3.69 (2H, s, H₂-7), 3.36 (2H, dt, *J*=5.9 and 6.8 Hz, H₂-10), 2.59 (2H, t, *J*=6.8 Hz, H₂-11); ^{13}C nmr (DMSO-*d*₆, 100 MHz) δ 163.4 (s, C-9), 151.5 (s, C-8), 149.2 (s, C-4), 147.0 (s, C-14), 132.5 (2C, d, C-2 and C-6), 131.4 (s, C-1), 124.5 (s, C-12), 111.9 (2C, s, C-3 and C-5), 109.4 (d, C-13), 37.5 (t, C-10), 27.7 (t, C-7), 24.6 (t, C-11); fabms [M+H]⁺ *m/z* 460, 462, 464 (1:2:1); hrfabms found *m/z* 459.9631, calcd for C₁₄H₁₃N₆O₃Br₂ [M+H]⁺ 459.9620.

ACKNOWLEDGMENTS

We thank Dr. T. Hoshino (Mukaishima Marine Biological Station, Hiroshima University) for identification of the sponge and Mr. Z. Nagahama for his help with collection of the sponge. This work was partly supported by a Grant-in-Aid from Ciba-Geigy Foundation, Japan, for the Promotion of Science and by a Grant-in-Aid for Scientific Research from the Ministry of Education, Science, and Culture of Japan.

LITERATURE CITED

1. J. Kobayashi and M. Ishibashi, in: "The Alkaloids." Ed. by A. Brossi and G.A. Cordell, Academic Press, San Diego, 1992, Vol. 41, p. 41.
2. H. Wu, H. Nakamura, J. Kobayashi, Y. Ohizumi, and Y. Hirata, *Experientia*, **42**, 855 (1986).
3. M. Tsuda, T. Sasaki, and J. Kobayashi, *J. Org. Chem.*, **59**, 3734 (1994).

4. J. Kobayashi, M. Tsuda, N. Kawasaki, K. Matsumoto, and T. Adachi, *Tetrahedron Lett.*, **35**, 4383 (1994).
5. M. Tsuda, N. Kawasaki, and J. Kobayashi, *Tetrahedron Lett.*, **35**, 4387 (1994).
6. H. Nakamura, H. Wu, J. Kobayashi, Y. Nakamura, Y. Ohizumi, and Y. Hirata, *Tetrahedron Lett.*, **26**, 4517 (1985).
7. L. Arabshahi and F.J. Schmitz, *J. Org. Chem.*, **52**, 3584 (1987).
8. F.W. MacLafferty, *Science*, **214**, 280 (1981).
9. M.L. Gross, *Int. J. Mass Spectrom. Ion Processes*, **118/119**, 137 (1992).
10. J.G. Batchelor, R.J. Cushley, and J.H. Prestegard, *J. Org. Chem.*, **39**, 1698 (1974).
11. A. Longeon, M. Guyot, and J. Vacelet, *Experientia*, **46**, 548 (1990).
12. T. Ichiba, P.J. Scheuer, and M. Kelly-Borges, *J. Org. Chem.*, **58**, 4149 (1993).
13. J. Jurek, W.Y. Yoshida, P.J. Scheuer, and M. Kelly-Borges, *J. Nat. Prod.*, **56**, 1609 (1993).

Received 5 October 1994